



Recent progress in photoacoustic molecular imaging

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By acoustically detecting the optical absorption contrast, photoacoustic (PA) tomography (PAT) has broken the penetration limits of traditional high-resolution optical imaging. Through spectroscopic analysis of the target's optical absorption, PAT can identify a wealth of endogenous and exogenous molecules and thus is inherently capable of molecular imaging with high sensitivity. PAT's molecular sensitivity is uniquely accompanied by non-ionizing radiation, high spatial resolution, and deep penetration in biological tissues, which other optical imaging modalities cannot achieve yet. In this concise review, we summarize the most recent technological advancements in PA molecular imaging and highlight the novel molecular probes specifically made for PAT in deep tissues. We conclude with a brief discussion of the opportunities for future advancements.

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Introduction

Photoacoustic tomography (PAT, also referred to as optoacoustic or thermoacoustic tomography) is based on the photoacoustic effect [1], in which ultrasonic waves generated by optical excitation are detected to map the original optical energy deposition [2–6]. PAT naturally utilizes rich optical absorption contrast and weak acoustic scattering inherent in biological tissue, lending it a clear advantage over traditional high-resolution optical imaging in retrieving anatomical, functional, molecular, metabolic, and histologic information at large depths.

One of the strengths of PAT is its inherent molecular sensitivity. Unlike fluorescent imaging which relies on the fluorescent molecule's radiative relaxation, PAT depends on the molecule's thermoelastic expansion through nonradiative relaxation. PAT has a 100% relative sensitivity to small optical absorption variations, meaning that a given percentage change in the optical absorption coefficient yields the same percentage change in the PA signal amplitude [7]. All molecules have unique optical absorption features that can serve as their 'fingerprints' for PA identification. The spatial distribution and optical properties of molecules are often closely related to their microenvironment (e.g. hypoxia in tumors), allowing PAT to probe physiological functions and pathological conditions. Moreover, because acoustic scattering in tissue is much weaker than optical scattering, PAT can harness scattered photons to explore molecular information with high spatial resolution at depths far beyond the optical diffusion limit (~1 mm).

PA molecular imaging has advanced rapidly since the first reports on PAT of the tumor microenvironment in the 2000s [8–10,11*,12]. Almost every technical aspect in PA molecular imaging has progressed, including the detection sensitivity and penetration depth of the imaging system [5,13–19,20**,21**,22*], the quantification accuracy of the signal unmixing [3,13,23–25], and the design and application of molecular probes in deep tissues [10,26–35,36**,37–39]. In this concise review, we focus on the major advancements in PA molecular imaging reported in the last several years (2014–2017) including novel imaging systems, signal unmixing methods, and molecular probes. We also overview the opportunities that may lead to future advances. Readers are referred to recent review articles to gain a more comprehensive knowledge of the principles of PAT [40,41], the molecular contrast agents [34,37,42,26,43], and the biomedical applications [6,16,44–46].

Basic principles of PAT

A typical PAT system includes a short-pulsed laser for efficient wideband PA signal generation, an ultrasonic transducer (or transducer array) for signal detection, a signal amplification and digitization system, and a computer for image formation. PAT has been implemented with two major image formation methods [2]. The first method, direct image formation, is based on mechanical scanning of a focused or unfocused excitation light beam and a focused single-element ultrasonic transducer. The second method, reconstruction image formation, is based on wide-field light illumination and parallel acoustic detection by a multi-element ultrasonic transducer array.

Direct image formation is commonly used in photoacoustic microscopy (PAM), whereas reconstruction image formation is the basis for photoacoustic computed tomography (PACT). Compared to PAM, PACT typically has a higher imaging speed and greater penetration depth but lower spatial resolutions [41]. Depending on the image formation method, PAT may require mechanical or electronic scanning to form two-dimensional (2D) and three-dimensional (3D) images.

PAT complements other imaging methods in contrast mechanism, spatial-temporal resolution, and penetration depth, and has found broad applications in the biomedical research, especially in functional brain mapping [47], cancer diagnosis and staging [44,48], tissue engineering and regenerative medicine [49], developmental biology [50], and molecular cell biology [51], as comprehensively reviewed elsewhere [6,52,53]. In particular, PAT has been widely used for various cancer studies [44], including fundamental research of cancerogenesis [54], cancer detection and staging [55], and navigation and evaluation in cancer treatment [56]. Using either endogenous contrast (e.g. melanin in melanoma cells) or exogenous contrast (e.g. targeted nanoparticles or organic dyes), PAT has become increasingly popular in providing accurate and early diagnosis of cancers [44].

Advances in PAT implementations

Continuous developments in laser technology, ultrasonic detection, digitization electronic systems, and parallel computation have driven technical breakthroughs in PAT technologies. Notably, inspired by PAT's rapid development and its increasingly important role in biomedical research, more and more manufacturers are developing commercial products specifically designed for PAT, including high-energy, high-speed pulsed lasers (e.g. pulsed laser diode illuminator, Quantel-Laser, Inc.), ultra-wideband ultrasonic transducers (e.g. 225 MHz bandwidth transducer, Olympus, Inc.), and high-speed, multi-channel data acquisition systems (e.g. 128 channel DAQ, Ultrasonix, Inc.). Industrial support in the development of PAT technology is critical for accelerating its commercialization and clinical translation.

Here, we highlight several recent technological breakthroughs in PAT. First, Real-time, whole-body small animal imaging has been achieved due to high-speed laser sources and data acquisition systems [20**,21**]. We reported a panoramic PACT system with a 125 μm in-plane resolution, 50 Hz 2D frame rate, and 48 mm penetration depth, which is capable of capturing circulating tumor cells in mouse brains (Figure 1a) [20**]. Fehm *et al.* developed a 3D PACT system to capture the dynamics of an entire heart beat with a 100 Hz 3D frame rate within a 1.5 cm^3 volume (Figure 1b) [21**]. These real-time, whole-body PAT systems are extremely powerful when tracking exogenously labeled drug molecules

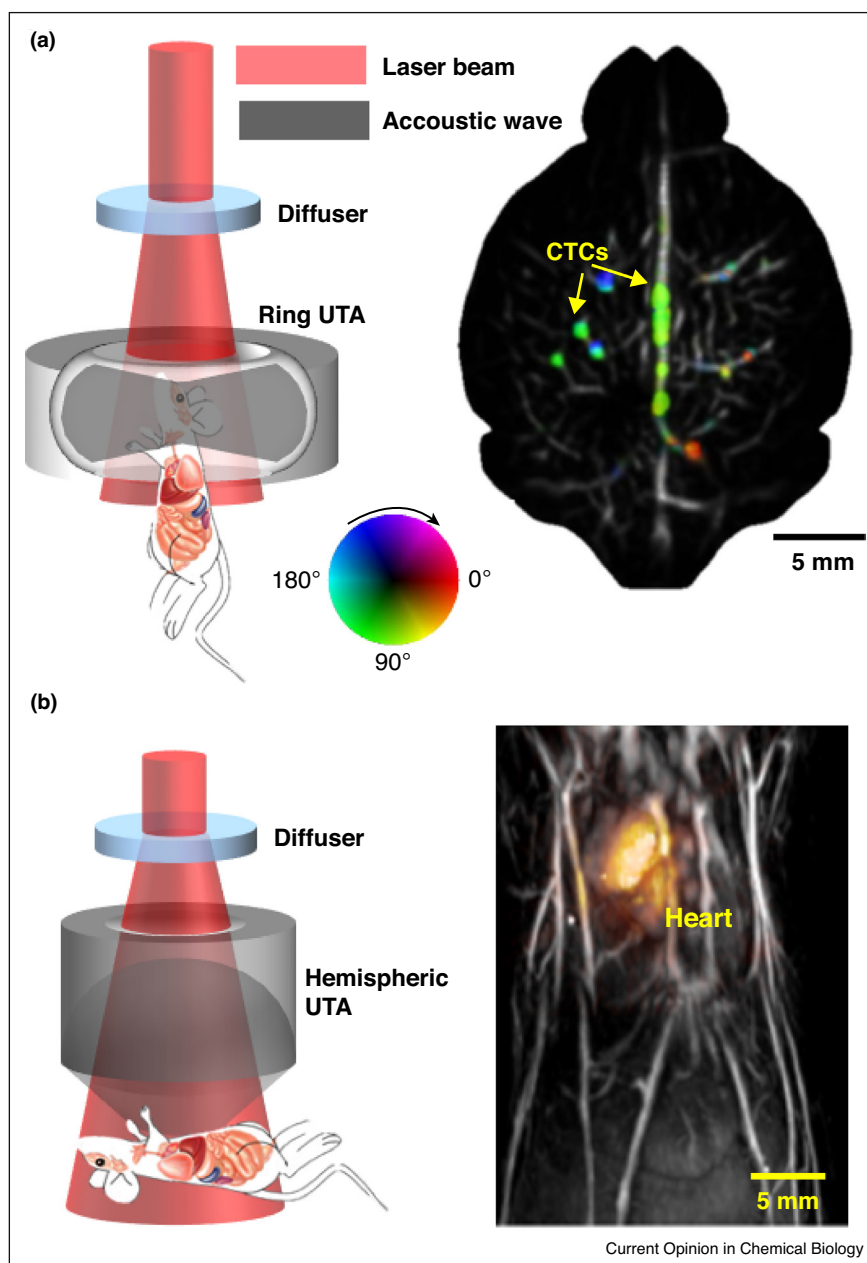
in pharmacokinetic studies on small animal models, thereby enabling biomedical researchers to test new drugs and monitor longitudinal therapy in the future. Second, The spatial resolutions of PAT have been pushing the existing limits through the use of high-frequency wideband ultrasound detection [22*,57*]. Aguirre *et al.* recently reported an ultra-broadband PAM system for human skin imaging with a spatial resolution of 7 μm , enabled by an ultrasonic detection band of 10–180 MHz [22*]. Guggenheim *et al.* developed a PAM system using an ultrasensitive plano-concave microresonator with an ultrasound detection band of 0–40 MHz and a large acceptance angle of 75 degrees [57*]. By matching the ultrasonic detection band with the detectable PA signal spectrum, which is primarily limited by the depth of the target, ultra-wideband ultrasonic detection has enabled multi-scale PA molecular imaging with the highest possible resolutions at different depths.

Advances in signal unmixing methods

Traditionally, spectroscopic imaging is used in PAT to extract the weak signals of molecular probes from the strong background signals of blood, by taking measurements at multiple optical wavelengths [3]. However, this method performs optimally only in superficial tissue because it requires knowledge of the local optical fluence (J/m^2), which is difficult to estimate in deep tissue [13,25]. Novel methods based on two different strategies have been developed to improve the signal unmixing accuracy in PAT [3,16,18,23,25]. The first strategy focuses on optical fluence compensation with tissue property modeling [18,23,24,58–71]. Instead of assuming homogenous optical properties, these advanced model-based methods typically treat the wavelength-dependent local optical fluence as another unknown parameter and then iteratively solve for the concentrations of molecular probes as an inverse problem. For example, Tzoumas *et al.* have recently reported an eigenspectrum-based method that has shown improved accuracy in quantifying deep-tissue blood oxygenation. This method models the local optical fluence as an affine function of only three reference base spectra (Figure 2a,b) [72**]. While this strategy has the potential to recover the concentrations of weakly-absorbing molecular probes, a large number of optical wavelengths (or reference fluence spectra) are needed, which slows down image acquisition. Moreover, the inverse problem is typically ill-posed and computationally intensive.

The second strategy seeks to recover the signal contribution from the molecular probes by exploring the temporal changes of the detected signals, assuming that the changes are confined only to the local molecular probes of interest [27,73–81,82**]. The temporal signal changes can be induced externally (e.g. light illumination) [36**,73] or internally (e.g. chemical cleavage) [74,75,82**]. For

Figure 1

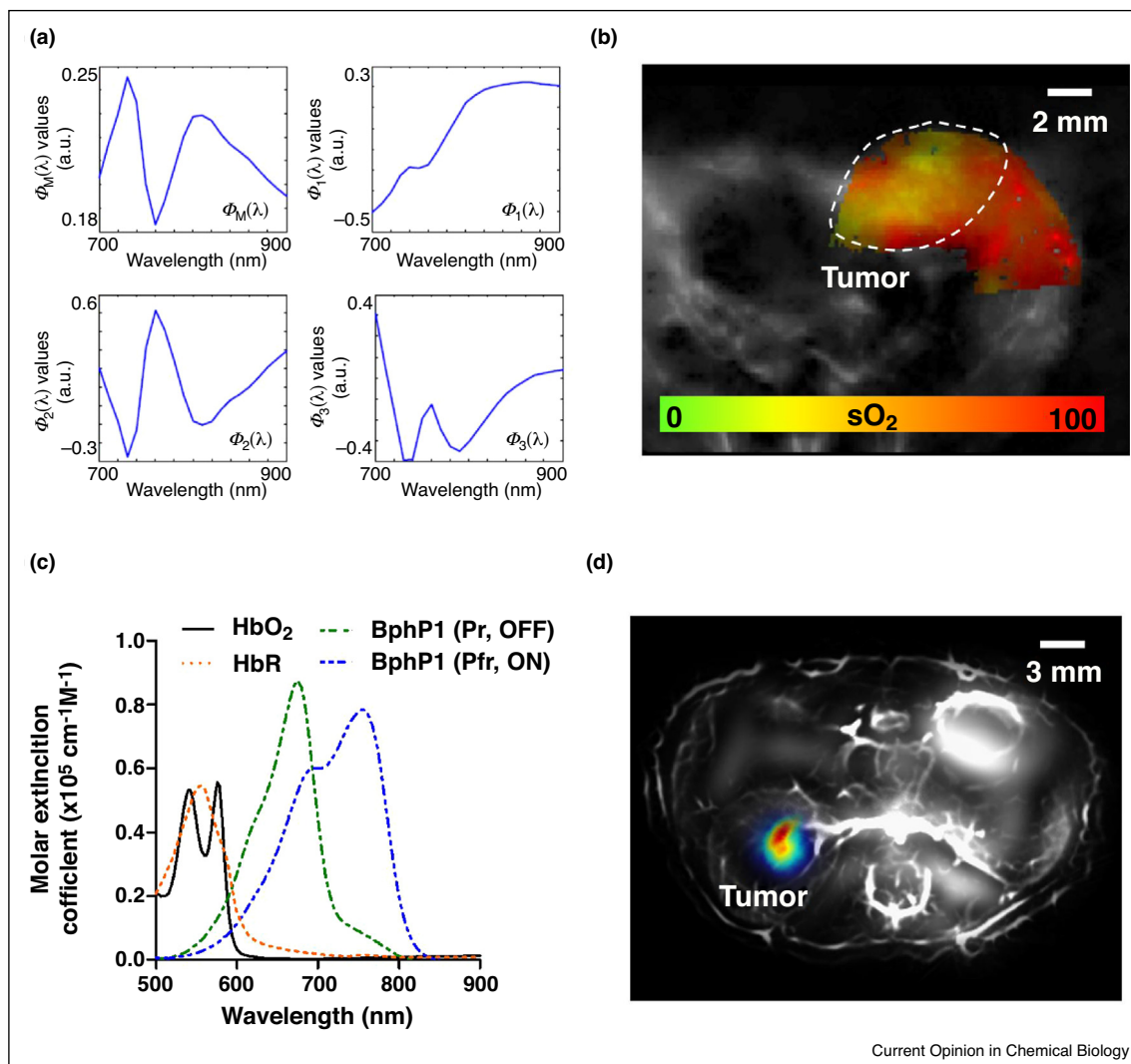


Advances in real-time, whole-body small animal PAT. **(a)** A ring-shaped ultrasonic transducer array (UTA) based panoramic PACT system with a 50 Hz 2D frame rate and 48 mm penetration depth, which is capable of capturing circulating tumor cells (CTCs) in mouse brains [20^{**}]. The colors represent the flow direction of CTCs. Flow speed is radially encoded in the color disk by hue saturation (a greater radius indicates a faster flow speed). **(b)** A hemispherical-shaped UTA based PACT system with a 100 Hz 3D frame rate in a 1.5 cm³ volume, which is capable of capturing the mouse heart beating [21^{**}].

example, several groups (including the authors') have explored the reversible photoswitching capability of several fluorescent (Dronpa, rsTagRFP) and non-fluorescent (BphP1, AGP1) proteins [36^{**},73,83]. By turning the molecular probe's optical absorption on or off at a certain wavelength, this temporal modulation can effectively eliminate the constant background signals without

needing to know the local optical fluence, thus dramatically enhance the image reconstruction robustness and detection sensitivity (Figure 2c,d). However, the applicability of this strategy is limited to special types of molecular probes whose optical properties can be physically or chemically modulated, such as activatable nanoparticles or photoswitchable proteins [27,36^{**}].

Figure 2



Advances in PAT signal unmixing. **(a)** An eigenspectrum-based signal unmixing method assumes the local optical fluence in deep tissue can be modeled as an affine function of three reference base spectra (Φ_1 , Φ_2 , and Φ_3) [72*]. **(b)** Eigenspectrum-based blood oxygenation mapping of the breast tumor in a mouse, showing the hypoxia tumor core. **(c)** A photoswitching-based signal unmixing method explores the two absorbing spectra of non-fluorescent protein BphP1. The constant background signals from hemoglobin can be suppressed through differential imaging [36*]. **(d)** Photoswitching-based differential image of the BphP1-expressing tumor in a mouse kidney.

Advances in molecular probes made for PAT

PAT does not rely on fluorescence emission of molecules, giving it the ability to image nearly all molecules, fluorescent or not. Taking advantage of wavelength-tunable optical parametric oscillator (OPO) lasers and Ti:Sapphire lasers, PAT has been implemented to explore numerous molecular probes with primary absorption wavelengths ranging from the ultraviolet to the near-infrared (NIR) region [26]. The ideal molecular probe for PAT should have the following attributes: be specific to the biological process of interest; exhibit maximal absorption in the NIR window for deep *in vivo* imaging; have zero or low fluorescent quantum yield (not strict); be nontoxic

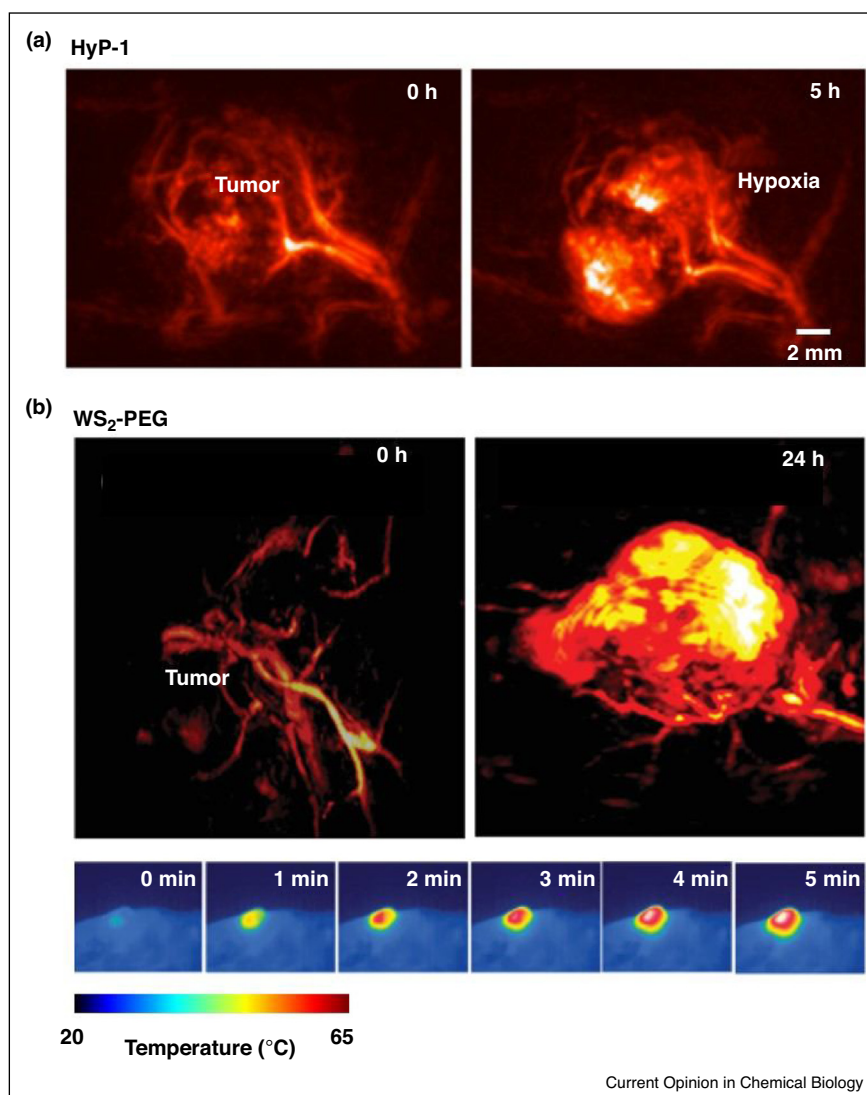
to the cells; and be resistant to photobleaching. As PAT draws increasing attention from the biomedical community at large, more and more molecular probes are being developed specifically for PAT by researchers in synthetic chemistry, protein engineering, and nanotechnology. The availability of commercial PAT systems has also accelerated the adoption of these molecular probes in fundamental research areas, including cancer biology [8,31,33,35,39,44,84,85], neuroscience [86,87*,88,89], and regenerative medicine [90–93].

So far, three major strategies have been individually or concurrently implemented in developing PAT-specific

molecular probes. First, Aiming to maximize the penetration depth of PAT, the first strategy focuses on developing contrast agents that have strong optical absorption in the NIR wavelength range with low fluorescent quantum yield [16,85,94–99,100**]. Taking advantage of melanin's strong absorption in the NIR range, Jathoul *et al.* developed a tyrosinase reporter gene system that introduced the key enzyme in melanin synthesis into non-melanogenic cells [100**]. Although melanin's relatively featureless absorption spectrum could make it hard to distinguish from intrinsic signals from hemoglobin, *in vivo* PAT of tyrosinase-expressing cells has shown high sensitivity [99,100**]. Zhou *et al.* recently developed a

phosphorus phthalocyanine (P-Pc) dye that has an absorption spectrum peaking around 1000 nm [98**]. P-Pc takes maximum advantage of its large molar extinction coefficient ($1.1 \times 10^5 \text{ cm}^{-1} \text{ M}^{-1}$ at 1064 nm) and the strong 1064 nm light from the Nd:YAG lasers, and thus has enabled deep tumor imaging *in vivo*. Second, Aiming to suppress the background signals from blood and improve the detection sensitivity, the second strategy focuses on developing contrast agents that can change their optical absorption in response to external or internal modulations [27,34,44,74–81,101]. Knox *et al.* reported an NIR agent for PA imaging of tissue hypoxia, which features an N-oxide-based trigger that can undergo facile

Figure 3



Advances in PAT molecular probes. (a) PACT images showing that HyP-1, an NIR hypoxia-response dye, changed its absorption peak from 670 nm to 760 nm five hours after exposure to the hypoxic environment of a breast tumor in a mouse [82**]. (b) PACT images of a mouse breast tumor before and 24 hours after i.v. injection of WS₂-PEG nanosheets, showing the accumulation of WS₂-PEG in the tumor region [102]. The bottom-row images show that, when exposed to 808 nm light, the photothermal effect of WS₂-PEG increased the local tumor temperature by 40 °C within 5 min.

bioreduction in the absence of oxygen and shifts the optical absorption peak from 670 nm to 760 nm (Figure 3a) [82^{••}]. By taking ratiometric measurement, the hypoxic tissue environment (e.g. tumor and ischemia) can be imaged. Third, Aiming to improve the theranostic efficiency in personalized medicine, the third strategy focuses on developing contrast agents that have simultaneous functionalities of imaging and therapy (e.g. photothermal, photodynamic, drug delivery) [77,81,102–105]. Cheng *et al.* demonstrated PEGylated nanosheets for dual-modal CT/PAT guided photothermal therapy of tumors [102]. The strong NIR absorption of the nanosheets provides excellent signals for PAT of the tumor structure, and efficient heating for ablating the tumor cells (Figure 3b).

Conclusion and discussions

Enabled by the advances in system implementations, signal unmixing methods, and molecular probes, PA molecular imaging has become increasingly popular in fundamental research and precision medicine. While this concise review can only cover a small portion of the exciting developments in PA molecular imaging, it has demonstrated the strong potential of this promising technology to continue growing and developing. It is also clear that the development of PAT has become a multidisciplinary effort from laser technology, ultrasound detection, high-speed electronics, mathematics, parallel computation, synthetic chemistry, protein engineering, and nanotechnology. The rapid growth of PAT technologies and their broad applications in biomedical research have, in turn, triggered new opportunities for each discipline.

With a series of long-standing engineering challenges overcome, we believe that PA molecular imaging will see even faster growth in the coming years. In particular, we anticipate four key breakthroughs. First, PA molecular imaging at depths around the optical dissipation limit (~10 cm) will be possible by developing molecular probes that can strongly absorb light in the NIR optical window, while other intrinsic tissue components present the least optical attenuations [106–108]. For example, the effective attenuation coefficient spectrum of human breast tissue has a minimum near 730 nm. Moreover, when the optical scattering effect is compensated for by using wavefront engineering technologies [109,110], PA molecular imaging may approach a sufficient penetration beyond 10 cm. Second, Single-molecule detection by PAT is highly promising using novel ultrasonic detectors with high piezoelectric efficiency (for piezoelectric ultrasound receivers) or high *Q*-factors (for optical ultrasound receivers) [57[•],111,112]. Third, Quantitative PA molecular imaging with high accuracy at depths will be enabled by new imaging methods and mathematical models that can better map the optical properties of the tissue [72^{••}]. Fourth, Finally, PA molecular imaging of neural activities in the deep brain will be achieved by using novel

genetically encodable indicators of action potentials or surrogates (e.g. voltage-sensitive or calcium-sensitive proteins) with strong absorption in the NIR spectral region [113].

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References and recommended reading

Papers of particular interest, published within the period of review, have been highlighted as:

- of special interest
- of outstanding interest

1. Bell AG: **Upon the production and reproduction of sound by light.** *Am J Sci* 1880, **20**:305-324.
2. Wang LV, Hu S: **Photoacoustic tomography: in vivo imaging from organelles to organs.** *Science* 2012, **335**:1458-1462.
3. Cox B, Laufer JG, Arridge SR, Beard PC: **Quantitative spectroscopic photoacoustic imaging: a review.** *J Biomed Opt* 2012, **17**:061202.
4. Greneur CL, Sagot B: *Biomedical Photoacoustic Imaging Patent Landscape.* 2015.
5. Taruttis A, Ntziachristos V: **Advances in real-time multispectral optoacoustic imaging and its applications.** *Nat Photon* 2015, **9**:219-227.
6. Zackrisson S, van de Ven SMWY, Gambhir SS: **Light in and sound out: emerging translational strategies for photoacoustic imaging.** *Cancer Res* 2014, **74**:979-1004.
7. Wang LV: **Tutorial on photoacoustic microscopy and computed tomography.** *IEEE J Sel Top Quantum Electron* 2008, **14**:171-179.
8. Copland JA, Eghtedari M, Popov VL, Kotov N, Mamedova N, Motamedi M, Oraevsky AA: **Bioconjugated gold nanoparticles as a molecular based contrast agent: implications for imaging of deep tumors using optoacoustic tomography.** *Mol Imaging Biol* 2004, **6**:341-349.
9. Mallidi S, Larson T, Aaron J, Sokolov K, Emelianov S: **Molecular specific optoacoustic imaging with plasmonic nanoparticles.** *Opt Express* 2007, **15**:6583-6588.
10. De La Zerda A, Zavaleta C, Keren S, Vaithilingam S, Bodapati S, Liu Z, Levi J, Smith BR, Ma TJ, Oralkan O, Cheng Z, Chen XY, Dai HJ, Khuri-Yakub BT, Gambhir SS: **Carbon nanotubes as photoacoustic molecular imaging agents in living mice.** *Nat Nanotechnol* 2008, **3**:557-562.
11. Li ML, Oh JT, Xie XY, Ku G, Wang W, Li C, Lungu G, Stoica G, Wang LV: **Simultaneous molecular and hypoxia imaging of brain tumors in vivo using spectroscopic photoacoustic tomography.** *Proc IEEE* 2008, **96**:481-489.
- The first demonstration of PA molecular imaging *in vivo*.
12. Li PC, Wang CRC, Shieh DB, Wei CW, Liao CK, Poe C, Jhan S, Ding AA, Wu YN: **In vivo photoacoustic molecular imaging with simultaneous multiple selective targeting using antibody-conjugated gold nanorods.** *Opt Express* 2008, **16**:18605-18615.
13. Razansky D, Baeten J, Ntziachristos V: **Sensitivity of molecular target detection by multispectral photoacoustic tomography (MSOT).** *Med Phys* 2009, **36**:939-945.

14. Razansky D, Distel M, Vinegoni C, Ma R, Perrimon N, Koster RW, Ntziachristos V: **Multispectral opto-acoustic tomography of deep-seated fluorescent proteins in vivo.** *Nat Photon* 2009, **3**:412-417.
15. Homan K, Kim S, Chen YS, Wang B, Mallidi S, Emelianov S: **Prospects of molecular photoacoustic imaging at 1064 nm wavelength.** *Opt Lett* 2010, **35**:2663-2665.
16. Razansky D, Deliolanis NC, Vinegoni C, Ntziachristos V: **Deep tissue optical and optoacoustic molecular imaging technologies for pre-clinical research and drug discovery.** *Curr Pharm Biotechnol* 2012, **13**:504-522.
17. Wang P, Rajian JR, Cheng JX: **Spectroscopic imaging of deep tissue through photoacoustic detection of molecular vibration.** *J Phys Chem Lett* 2013, **4**:2177-2185.
18. Tzoumas S, Nunes A, Deliolanis NC, Ntziachristos V: **Effects of multispectral excitation on the sensitivity of molecular optoacoustic imaging.** *J Biophoton* 2015, **8**:629-637.
19. Hui J, Li R, Phillips EH, Goergen CJ, Sturek M, Cheng JX: **Bond-selective photoacoustic imaging by converting molecular vibration into acoustic waves.** *Photoacoustics* 2016, **4**:11-21.
20. Li L, Zhu L, Ma C, Lin L, Yao J, Wang L, Maslov K, Zhang R, Chen W, Shi J, Wang LV: **Single-impulse panoramic photoacoustic computed tomography of small-animal whole-body dynamics at high spatiotemporal resolution.** *Nat Biomed Eng* 2017, **1**:0071.
- The first panoramic PACT system that can provide real-time imaging of small-animal whole-body dynamics, with submillimeter resolutions. The ring-shaped 512-elements ultrasonic transducer array surrounded the animal and covered the entire cross-sectional region.
21. Fehm TF, Dean-Ben XL, Ford SJ, Razansky D: **In vivo whole-body optoacoustic scanner with real-time volumetric imaging capacity.** *Optica* 2016, **3**:1153-1159.
- The first demonstration of 3D PACT with real-time imaging of small-animal regional dynamics. The semi-spherical 256-elements ultrasonic transducer array provided near-isotropic spatial resolutions over a 1.5 cm³ tissue volume. However, 256 elements are considered too sparse for a 2D array, tending to cause imaging artifacts outside the central zone of the 3D field of view covered by the array.
22. Aguirre J, Schwarz M, Garzorz N, Omar M, Buehler A, Eyerich K, Ntziachristos V: **Precision assessment of label-free psoriasis biomarkers with ultra-broadband optoacoustic mesoscopy.** *Nat Biomed Eng* 2017, **1**:0068.
- The utilization of an ultra-wideband ultrasonic transducer enabled high-resolution PAM of human skins at different depths.
23. Tzoumas S, Deliolanis NC, Morscher S, Ntziachristos V: **Unmixing molecular agents from absorbing tissue in multispectral optoacoustic tomography.** *IEEE Trans Med Imaging* 2014, **33**:48-60.
24. Tzoumas S, Kravtsov A, Gao Y, Buehler A, Ntziachristos V: **Statistical molecular target detection framework for multispectral optoacoustic tomography.** *IEEE Trans Med Imaging* 2016, **35**:2534-2545.
25. Beard PC, Lauffer JG, Cox B, Arridge SR: **Quantitative photoacoustic imaging: measurement of absolute chromophore concentrations for physiological and molecular imaging.** In *Photoacoustic Imaging and Spectroscopy*, vol 144. Edited by Wang LV. Boca Raton: CRC Press-Taylor & Francis Group; 2009:121-143.
26. Weber J, Beard PC, Bohndiek SE: **Contrast agents for molecular photoacoustic imaging.** *Nat Methods* 2016, **13**:639-650.
27. Levi J, Kothapalli SR, Ma TJ, Hartman K, Khuri-Yakub BT, Gambhir SS: **Design, synthesis, and imaging of an activatable photoacoustic probe.** *J Am Chem Soc* 2010, **132**:11264-11269.
28. Bayer CL, Chen YS, Kim S, Mallidi S, Sokolov K, Emelianov S: **Multiplex photoacoustic molecular imaging using targeted silica-coated gold nanorods.** *Biomed Opt Express* 2011, **2**:1828-1835.
29. de la Zerda A, Kim JW, Galanzha EI, Gambhir SS, Zharov VP: **Advanced contrast nanoagents for photoacoustic molecular imaging, cytometry, blood test and photothermal theranostics.** *Contrast Media Mol Imaging* 2011, **6**:346-369.
30. Pan D, Pramanik M, Wickline SA, Wang LHV, Lanza GM: **Recent advances in colloidal gold nanobeacons for molecular photoacoustic imaging.** *Contrast Media Mol Imaging* 2011, **6**:378-388.
31. Kircher MF, de la Zerda A, Jockerst JV, Zavaleta CL, Kempen PJ, Mittra E, Pitter K, Huang RM, Campos C, Habte F, Sinclair R, Brennan CW, Mellinghoff IK, Holland EC, Gambhir SS: **A brain tumor molecular imaging strategy using a new triple-modality MRI-photoacoustic-Raman nanoparticle.** *Nat Med* 2012, **18**:829-834.
32. Wei CW, Lombardo M, Larson-Smith K, Pelivanov I, Perez C, Xia J, Matula T, Pozzo D, O'Donnell M: **Nonlinear contrast enhancement in photoacoustic molecular imaging with gold nanosphere encapsulated nanoemulsions.** *Appl Phys Lett* 2014, **104**:4.
33. Balasundaram G, Ho CJH, Li K, Driessen W, Dinis US, Wong CL, Ntziachristos V, Liu B, Olivo M: **Molecular photoacoustic imaging of breast cancer using an actively targeted conjugated polymer.** *Int J Nanomed* 2015, **10**:387-397.
34. Miao QQ, Pu KY: **Emerging designs of activatable photoacoustic probes for molecular imaging.** *Bioconjug Chem* 2016, **27**:2808-2823.
35. Onoe S: **Development of molecular probes for spatio-temporal analysis of in vivo tumor with photoacoustic imaging.** *Yakugaku Zasshi J Pharm Soc Jpn* 2016, **136**:491-498.
36. Yao J, Kaberniuk AA, Li L, Shcherbakova DM, Zhang R, Wang L, Li G, Verkhusa VV, Wang LV: **Multiscale photoacoustic tomography using reversibly switchable bacterial phytochrome as a near-infrared photochromic probe.** *Nat Methods* 2016, **13**:67-73.
- The first demonstration of switchable non-fluorescent protein BphP1 for PA molecular imaging. The switching capability of BphP1 enabled suppression of background signals from hemoglobin and thus improved the detection sensitivity.
37. Liu LM, Qin H: **Photoacoustic molecular imaging with functional nanoparticles.** *J Innov Opt Health Sci* 2017, **10**:12.
38. Liu Y, Wang S, Ma Y, Lin J, Wang HY, Gu YQ, Chen XY, Huang P: **Ratiometric photoacoustic molecular imaging for methylmercury detection in living subjects.** *Adv Mater* 2017, **29**:5.
39. Wilson KE, Bachawal SV, Abou-Elkacem L, Jensen K, Machtaler S, Tian L, Willmann JK: **Spectroscopic photoacoustic molecular imaging of breast cancer using a B7-H3-targeted ICG contrast agent.** *Theranostics* 2017, **7**:1463-1476.
40. Beard P: **Biomedical photoacoustic imaging.** *Interface Focus* 2011, **1**:602-631.
41. Wang LV, Yao J: **A practical guide to photoacoustic tomography in the life sciences.** *Nat Methods* 2016, **13**:627-638.
42. Kim C, Favazza C, Wang LHV: **In vivo photoacoustic tomography of chemicals: high-resolution functional and molecular optical imaging at new depths.** *Chem Rev* 2010, **110**:2756-2782.
43. Gujrati V, Mishra A, Ntziachristos V: **Molecular imaging probes for multi-spectral optoacoustic tomography.** *Chem Commun* 2017, **53**:4653-4672.
44. Wilson KE, Wang TY, Willmann JK: **Acoustic and photoacoustic molecular imaging of cancer.** *J Nucl Med* 2013, **54**:1851-1854.
45. Taruttis A, Ntziachristos V: **Optoacoustic molecular imaging: methods and applications.** In *Advanced Biophotonics: Tissue Optical Sectioning*. Edited by Wang RK, Tuchin VV. Boca Raton: CRC Press-Taylor & Francis Group; 2014:449-474.
46. Liu YJ, Nie LM, Chen XY: **Photoacoustic molecular imaging: from multiscale biomedical applications towards early-stage theranostics.** *Trends Biotechnol* 2016, **34**:420-433.
47. Yao J, Wang LV: **Photoacoustic brain imaging: from microscopic to macroscopic scales.** *Neurophotonics* 2014, **1**:011003.
48. Mallidi S, Luke GP, Emelianov S: **Photoacoustic imaging in cancer detection, diagnosis, and treatment guidance.** *Trends Biotechnol* 2011, **29**:213-221.

49. Cai X, Zhang YS, Xia Y, Wang LV: **Photoacoustic microscopy in tissue engineering**. *Mater Today* 2013, **16**:67-77.
50. Ripoll J, Koberstein-Schwarz B, Ntziachristos V: **Unleashing optics and optoacoustics for developmental biology**. *Trends Biotechnol* 2015, **33**:679-691.
51. Strohm EM, Moore MJ, Koliou MC: **Single cell photoacoustic microscopy: a review**. *IEEE J Sel Top Quantum Electron* 2015, **PP**:1.
52. Wang LHV, Hu S: **Photoacoustic tomography: in vivo imaging from organelles to organs**. *Science* 2012, **335**:1458-1462.
53. Wang LV: **Multiscale photoacoustic microscopy and computed tomography**. *Nat Photon* 2009, **3**:503-509.
54. Herzog E, Taruttis A, Beziere N, Lutich AA, Razansky D, Ntziachristos V: **Optical imaging of cancer heterogeneity with multispectral optoacoustic tomography**. *Radiology* 2012, **263**:461-468.
55. Kim JW, Galanzha EI, Shashkov EV, Moon HM, Zharov VP: **Golden carbon nanotubes as multimodal photoacoustic and photothermal high-contrast molecular agents**. *Nat Nanotechnol* 2009, **4**:688-694.
56. Laufer J, Johnson P, Zhang E, Treeby B, Cox B, Pedley B, Beard P: **In vivo preclinical photoacoustic imaging of tumor vasculature development and therapy**. *J Biomed Opt* 2012, **17**:056016.
57. Guggenheim JA, Li J, Allen TJ, Colchester RJ, Noimark S, Ogunlade O, Parkin IP, Papakonstantinou I, Desjardins AE, Zhang EZ, Beard PC: **Ultrasensitive plano-concave optical microresonators for ultrasound sensing**. *Nat Photon* 2017, **11**:714-719.
- The development of an ultra-sensitive all-optical ultrasound detector for PA and ultrasound imaging, especially for endoscopic applications.
58. Cox BT, Laufer JG, Beard PC: **Quantitative photoacoustic image reconstruction using fluence dependent chromophores**. *Biomed Opt Express* 2010, **1**:201-208.
59. Yao L, Sun Y, Jiang HB: **Transport-based quantitative photoacoustic tomography: simulations and experiments**. *Phys Med Biol* 2010, **55**:1917-1934.
60. Zemp RJ: **Quantitative photoacoustic tomography with multiple optical sources**. *Appl Opt* 2010, **49**:3566-3572.
61. Bauer AQ, Nothdurft RE, Erpelding TN, Wang LHV, Culver JP: **Quantitative photoacoustic imaging: correcting for heterogeneous light fluence distributions using diffuse optical tomography**. *J Biomed Opt* 2011, **16**:096016.
62. Li SF, Montcel B, Liu WY, Vray D: **Analytical model of optical fluence inside multiple cylindrical inhomogeneities embedded in an otherwise homogeneous turbid medium for quantitative photoacoustic imaging**. *Opt Express* 2014, **22**:20500-20514.
63. Song NN, Deumic C, Da Silva A: **Considering sources and detectors distributions for quantitative photoacoustic tomography**. *Biomed Opt Express* 2014, **5**:3960-3974.
64. Malone E, Powell S, Cox BT, Arridge S: **Reconstruction-classification method for quantitative photoacoustic tomography**. *J Biomed Opt* 2015, **20**:126004.
65. Hochuli R, Powell S, Arridge S, Cox B: **Quantitative photoacoustic tomography using forward and adjoint Monte Carlo models of radiance**. *J Biomed Opt* 2016, **21**:126004.
66. Liu YB, Jiang HB, Yuan Z: **Two schemes for quantitative photoacoustic tomography based on Monte Carlo simulation**. *Med Phys* 2016, **43**:3987-3997.
67. Venugopal M, van Es P, Manohar S, Roy D, Vasu RM: **Quantitative photoacoustic tomography by stochastic search: direct recovery of the optical absorption field**. *Opt Lett* 2016, **41**:4202-4205.
68. An L, Saratoon T, Fonseca M, Ellwood R, Cox B: **Statistical independence in nonlinear model-based inversion for quantitative photoacoustic tomography**. *Biomed Opt Express* 2017, **8**:5297-5310.
69. Brochu FM, Brunker J, Joseph J, Tomaszewski MR, Morscher S, Bohndiek SE: **Towards quantitative evaluation of tissue absorption coefficients using light fluence correction in optoacoustic tomography**. *IEEE Trans Med Imaging* 2017, **36**:322-331.
70. Nykanen O, Pulkkinen A, Tarvainen T: **Quantitative photoacoustic tomography augmented with surface light measurements**. *Biomed Opt Express* 2017, **8**:4380-4395.
71. Shan TQ, Qi J, Jiang M, Jiang HB: **GPU-based acceleration and mesh optimization of finite-element-method-based quantitative photoacoustic tomography: a step towards clinical applications**. *Appl Opt* 2017, **56**:4426-4432.
72. Tzoumas S, Nunes A, Olefir I, Stangl S, Symvoulidis P, Glasl S, Bayer C, Multhoff G, Ntziachristos V: **Eigenspectra optoacoustic tomography achieves quantitative blood oxygenation imaging deep in tissues**. *Nat Commun* 2016, **7**:12121.
- This study reported a new mathematic method for quantifying the absorption coefficients in PA molecular imaging, in which the local optical fluence was modeled as a linear combination of three independent eigenspectra.
73. Stiel AC, Dean-Ben XL, Jiang YY, Ntziachristos V, Razansky D, Westmeyer GG: **High-contrast imaging of reversibly switchable fluorescent proteins via temporally unmixing multispectral optoacoustic tomography**. *Opt Lett* 2015, **40**:367-370.
74. Dragulescu-Andrasi A, Kothapalli SR, Tikhomirov GA, Rao JH, Gambhir SS: **Activatable oligomerizable imaging agents for photoacoustic imaging of furin-like activity in living subjects**. *J Am Chem Soc* 2013, **135**:11015-11022.
75. Hirasawa T, Iwatate RJ, Kamiya M, Okawa S, Urano Y, Ishihara M: **Multispectral photoacoustic imaging of tumours in mice injected with an enzyme-activatable photoacoustic probe**. *J Opt* 2017, **19**:14.
76. Jeon M, Song WT, Huynh E, Kim J, Kim J, Helfield BL, Leung BYC, Goertz DE, Zheng G, Oh J, Lovell JF, Kim C: **Methylene blue microbubbles as a model dual-modality contrast agent for ultrasound and activatable photoacoustic imaging**. *J Biomed Opt* 2014, **19**:8.
77. Liang XL, Fang L, Li XD, Zhang X, Wang F: **Activatable near infrared dye conjugated hyaluronic acid based nanoparticles as a targeted theranostic agent for enhanced fluorescence/CT/photoacoustic imaging guided photothermal therapy**. *Biomaterials* 2017, **132**:72-84.
78. Miao QQ, Lyu Y, Ding D, Pu KY: **Semiconducting oligomer nanoparticles as an activatable photoacoustic probe with amplified brightness for in vivo imaging of pH**. *Adv Mater* 2016, **28**:3662-3668.
79. Morgounova E, Shao Q, Hackel BJ, Thomas DD, Ashkenazi S: **Photoacoustic lifetime contrast between methylene blue monomers and self-quenched dimers as a model for dual-labeled activatable probes**. *J Biomed Opt* 2013, **18**:8.
80. Zhang JJ, Zhen X, Upputuri PK, Pramanik M, Chen P, Pu KY: **Activatable photoacoustic nanoprobe for in vivo ratiometric imaging of peroxynitrite**. *Adv Mater* 2017, **29**:8.
81. Zhang LW, Gao S, Zhang F, Yang K, Ma QJ, Zhu L: **Activatable hyaluronic acid nanoparticle as a theranostic agent for optical/photoacoustic image-guided photothermal therapy**. *ACS Nano* 2014, **8**:12250-12258.
82. Knox HJ, Hedhli J, Kim TW, Khalili K, Dobrucki LW, Chan J: **A bioreducible N-oxide-based probe for photoacoustic imaging of hypoxia**. *Nat Commun* 2017, **8**:1794.
- This study developed the first near-infrared PA probe Hyp-1 that could report the local hypoxic microenvironment. In a low-oxygen environment, Hyp-1 can be converted into red-Hyp-1 with a different optical absorption spectrum.
83. Dortay H, Märk J, Wagener A, Zhang E, Grötzinger C, Hildebrandt P, Friedrich T, Laufer J: **Dual-wavelength photoacoustic imaging of a photoswitchable reporter protein**. *SPIE BIOS*. SPIE; 2016.
84. Sano K: **Development of molecular probes based on iron oxide nanoparticles for in vivo magnetic resonance/photoacoustic**

- dual imaging of target molecules in tumors.** *Yakugaku Zasshi J Pharm Soc Jpn* 2017, **137**:55-60.
85. Xiang LZ, Yuan Y, Xing D, Ou ZM, Yang SH, Zhou FF: **Photoacoustic molecular imaging with antibody-functionalized single-walled carbon nanotubes for early diagnosis of tumor.** *J Biomed Opt* 2009, **14**:7.
86. Dean-Ben XL, Gottschalk S, Sela G, Shoham S, Razansky D: **Functional optoacoustic neuro-tomography of calcium fluxes in adult zebrafish brain in vivo.** *Opt Lett* 2017, **42**:959-962.
87. Dean-Ben XL, Sela G, Lauri A, Kneipp M, Ntziachristos V, Westmeyer GG, Shoham S, Razansky D: **Functional optoacoustic neuro-tomography for scalable whole-brain monitoring of calcium indicators.** *Light Sci Appl* 2016, **5**:7.
- The first report of PA imaging of neural activities *in vivo*, using a fluorescent calcium-sensitive indicator GCaMP. However, using a single wavelength of 488 nm, it is likely that the detected PA signal changes included the contributions from the concurrent hemodynamic responses.
88. Rasheed N, Cressman JR, Chitnis PV: **Feasibility of using RH795 dye for photoacoustic imaging of neuro-electrical activity.** In *Neural Imaging and Sensing*. Edited by Luo Q, Ding J. Bellingham: SPIE-Int Soc Optical Engineering; 2017.
89. Zhang RY, Rao B, Rong HY, Raman B, Wang LV: **In vivo photoacoustic neuronal imaging of odor-evoked calcium signals in the Drosophila brain (conference presentation).** In *Photons Plus Ultrasound: Imaging and Sensing 2016*. Edited by Oraevsky AA, Wang LV. Bellingham: SPIE-Int Soc Optical Engineering; 2016.
90. Avigo C, Flori A, Armanetti P, Di Lascio N, Kusmic C, Jose J, Losi P, Soldani G, Faïta F, Menichetti L: **Strategies for non-invasive imaging of polymeric biomaterial in vascular tissue engineering and regenerative medicine using ultrasound and photoacoustic techniques.** *Polym Int* 2016, **65**:734-740.
91. Ishihara M, Sato M, Sato S, Kikuchi T, Mitani G, Kaneshiro N, Ishihara M, Mochida J, Kikuchi M: **Usefulness of the photoacoustic measurement method for monitoring the regenerative process of full-thickness defects in articular cartilage using tissue-engineering technology.** In *Optical Interactions with Tissue and Cells XVI*. Edited by Jacques SL, Wang LV. Bellingham: SPIE-Int Soc Optical Engineering; 2005: 208-219.
92. Nam SY, Chung E, Suggs LJ, Emelianov SY: **Combined ultrasound and photoacoustic imaging to noninvasively assess burn injury and selectively monitor a regenerative tissue-engineered construct.** *Tissue Eng Part C Methods* 2015, **21**:557-566.
93. Sharkey J, Scarfe L, Comenge J, Brillant N, Burton N, Antoine D, Wilm B, Levy R, Park K, Murray P: **Utilising multispectral optoacoustic tomography (MSOT) to assess organ function and track labelled therapeutic cells for regenerative medicine therapies in vivo.** *Hum Gene Ther* 2017, **28**:A19.
94. Nie LM, Chen M, Sun XL, Rong PF, Zheng NF, Chen XY: **Palladium nanosheets as highly stable and effective contrast agents for in vivo photoacoustic molecular imaging.** *Nanoscale* 2014, **6**:1271-1276.
95. Pu KY, Shuhendler AJ, Jokerst JV, Mei JG, Gambhir SS, Bao ZN, Rao JH: **Semiconducting polymer nanoparticles as photoacoustic molecular imaging probes in living mice.** *Nat Nanotechnol* 2014, **9**:233-239.
96. Stahl T, Bofinger R, Lam I, Fallon KJ, Johnson P, Ogunlade O, Vassileva V, Pedley RB, Beard PC, Hailes HC, Bronstein H, Tabor AB: **Tunable semiconducting polymer nanoparticles with INDT-based conjugated polymers for photoacoustic molecular imaging.** *Bioconj Chem* 2017, **28**:1734-1740.
97. Wang HN, Liu CB, Gong XJ, Hu DH, Lin RQ, Sheng ZH, Zheng CF, Yan M, Chen JQ, Cai LT, Song L: **In vivo photoacoustic molecular imaging of breast carcinoma with folate receptor-targeted indocyanine green nanoprobe.** *Nanoscale* 2014, **6**:14270-14279.
98. Zhou Y, Wang DP, Zhang YM, Chitgupi U, Geng JM, Wang YH, Zhang YZ, Cook TR, Xia J, Lovell JF: **A phosphorus phthalocyanine formulation with intense absorbance at 1000 nm for deep optical imaging.** *Theranostics* 2016, **6**:688-697.
- A PA molecular probe that absorbs strongly at 1064 nm, which is the fundamental wavelength of the Nd:YAG lasers widely used in PAT systems.
99. Krumholz A, VanVickle-Chavez SJ, Yao JJ, Fleming TP, Gillanders WE, Wang LHV: **Photoacoustic microscopy of tyrosinase reporter gene in vivo.** *J Biomed Opt* 2011, **16**:080503.
100. Jathoul AP, Laufer J, Ogunlade O, Treeby B, Cox B, Zhang E, Johnson P, Pizzey AR, Philip B, Marafioti T, Lythgoe MF, Pedley RB, Pule MA, Beard P: **Deep in vivo photoacoustic imaging of mammalian tissues using a tyrosinase-based genetic reporter.** *Nat Photon* 2015, **9**:239-246.
- The first demonstrations of high-sensitivity PAT of nonmelanogenic mammalian tissues that transiently (Ref. [99]) and stably (Ref. [100**]) expressed melanin, respectively, using a tyrosinase-based gene reporter.
101. Levi J, Kothapalli SR, Bohndiek S, Yoon JK, Dragulescu-Andrasi A, Nielsen C, Tisma A, Bodapati S, Gowrishankar G, Yan XR, Chan C, Starcevic D, Gambhir SS: **Molecular photoacoustic imaging of follicular thyroid carcinoma.** *Clin Cancer Res* 2013, **19**:1494-1502.
102. Cheng L, Liu JJ, Gu X, Gong H, Shi XZ, Liu T, Wang C, Wang XY, Liu G, Xing HY, Bu WB, Sun BQ, Liu Z: **PEGylated WS2 nanosheets as a multifunctional theranostic agent for in vivo dual-modal CT/photoacoustic imaging guided photothermal therapy.** *Adv Mater* 2014, **26**:1886-1893.
103. Emelianov S, Mallidi S, Larson T, Sokolov K: **Photoacoustic imaging and therapy utilizing molecular specific plasmonic nanoparticles.** In *Photoacoustic Imaging and Spectroscopy*, vol 144. Edited by Wang LV. Boca Raton: CRC Press-Taylor & Francis Group; 2009:399-407.
104. Lyu Y, Fang Y, Miao QQ, Zhen X, Ding D, Pu KY: **Intraparticle molecular orbital engineering of semiconducting polymer nanoparticles as amplified theranostics for in vivo photoacoustic imaging and photothermal therapy.** *ACS Nano* 2016, **10**:4472-4481.
105. Wang B, Su J, Karpouk A, Yeager D, Emelianov S: **Intravascular photoacoustic and ultrasound imaging: from tissue characterization to molecular imaging to image-guided therapy.** In *Atherosclerosis Disease Management*. Edited by Suri JS, Kathuria C, Molinari F. New York: Springer; 2011:787-816.
106. Sordillo LA, Pu Y, Pratavieira S, Budansky Y, Alfano RR: **Deep optical imaging of tissue using the second and third near-infrared spectral windows.** *J Biomed Opt* 2014, **19**:056004.
107. Wilson RH, Nadeau KP, Jaworski FB, Tromberg BJ, Durkin AJ: **Review of short-wave infrared spectroscopy and imaging methods for biological tissue characterization.** *J Biomed Opt* 2015, **20**:030901.
108. Bashkatov AN, Genina EA, Kochubey VI, Tuchin VV: **Optical properties of human skin, subcutaneous and mucous tissues in the wavelength range from 400 to 2000 nm.** *J Phys D Appl Phys* 2005, **38**:2543-2555.
109. Horstmeyer R, Ruan HW, Yang CH: **Guidestar-assisted wavefront-shaping methods for focusing light into biological tissue.** *Nat Photon* 2015, **9**:563-571.
110. Lai P, Wang L, Tay JW, Wang LV: **Photoacoustically guided wavefront shaping for enhanced optical focusing in scattering media.** *Nat Photon* 2015, **9**:126-132.
111. Dong BQ, Chen SY, Zhang Z, Sun C, Zhang HF: **Photoacoustic probe using a microring resonator ultrasonic sensor for endoscopic applications.** *Opt Lett* 2014, **39**:4372-4375.
112. Xie ZX, Tian C, Chen SL, Ling T, Zhang C, Guo LJ, Carson PL, Wang XD: **3D high resolution photoacoustic imaging based on pure optical photoacoustic microscopy with microring resonator.** In *Photons Plus Ultrasound: Imaging and Sensing 2014*. Edited by Oraevsky AA, Wang LV. 2014.
113. Miller EW: **Small molecule fluorescent voltage indicators for studying membrane potential.** *Curr Opin Chem Biol* 2016, **33**:74-80.